

# AG23166\*A

Certificate of Analysis

Product Description	Transgenic Murine Embryonic Stem (mES) Cell containing the transcription factor Zscan4
Publication	Nishiyama et al.; PMID <u>19796622</u>
Passage of mES reported at submission	26
Number of passages at Coriell	3
Freeze Passage	29
Media	DMEM + 20% ES cell FBS + puromycin +
	doxycycline + LIF
Feeder	DR4 MEFs on 0.1% gelatin
Passage method	Accutase
Split ratio	Seed at 1.2 x 10 <sup>6</sup> cells per 1 well of 6 well plate
	$(1.0 \times 10^5 \text{ cells/cm}^2) \text{ split at } 80\% \text{ confluence } (2-3)$
	days)

#### The following testing specifications have been met for the specified product lot:

Test Description	Test Method	Test Specification	Result
Test Description	1 est Memou	rest opecification	Kesuit
Viability	Cell Count Post Thaw	Cells double within 3	
	of Cryopreserved	days after recovery	
	Cells		Pass
Sterility	Growth on agar	Negative	
			Pass
Mycoplasma	PCR	Negative	
			Pass
Karyotype	G-banding	At least 60% normal	
20 AND MOTOR	41	cells	94% 40 XY
Identity	Nucleoside	Murine	
	Phosphorylase		
	Isoenzyme		
	Electrophoresis	> 80% expression of	Pass
Surface Antigen	rface Antigen Immunostaining		
Expression		SSEA1	
			Pass
Pluripotency	Embryoid Body	Morphology and	
	Formation	expression of lineage-	
		specific genes	Pass
<b>Transgene Induction</b>	Doxycycline removal	Increase in transgene	
		expression by qPCR	8 Fold Change

### **Post-Thaw Viability**

One vial of was thawed after cryopreservation. Cells are counted following recovery and plated in one well of a 6 well plate. Cultures are observed daily and passaged when cells are approximately 80% confluent. Following dissociation with accutase, cells are counted and viable cell number is determined. The viable cell number must double within 3 days following recovery.

Days from Recovery to First	Viable Cell Number	Viable Cell Number at
Passage	at Thaw	First Passage
2	$2.63 \times 10^6$	$6.91 \times 10^6$

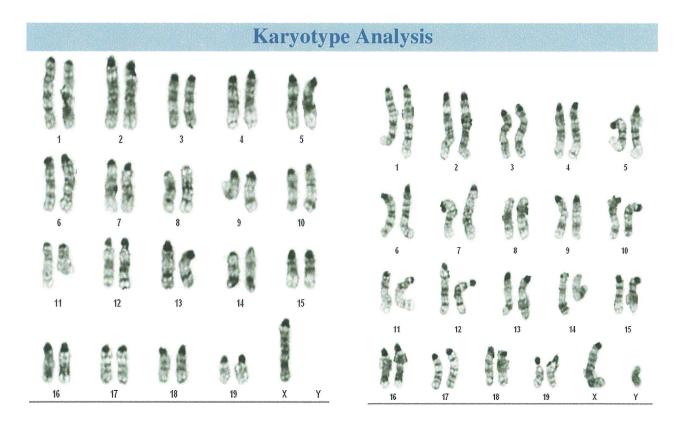


Figure 1A: Karyotype Image showing 39X.

Figure 1B: Karyotype Image showing 40XY.

## **Surface Antigen Expression of Stem Cell Markers**

Undifferentiated cells are stained for the surface antigens, SSEA1. SSEA1 (stage specific embryonic antigen 1) is expressed on undifferentiated murine stem cells.

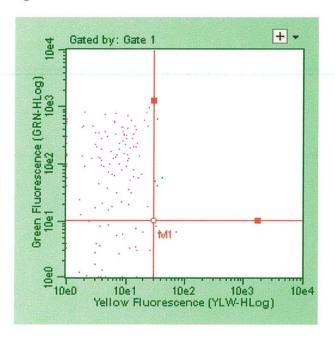


Figure 2A: Scatter plot of SSEA1 stained iPS cells.

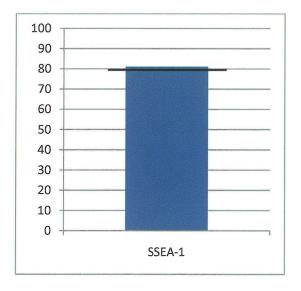


Figure 2B. Graph depicting percent SSEA1 positive cells in undifferentiated cell culture

### **Assessment of Pluripotency of a Cell Line**

Cells are subjected to direct differentiaion to assess the pluripotency of the cell line. RNA is harvested and gene expression is analyzed by quantitative real-time PCR. Ct values are adjusted for loading using a housekeeping gene. Gene expression is shown as fold difference to undifferentiated cells.

## **Embryoid Body (EB) Formation Assay**

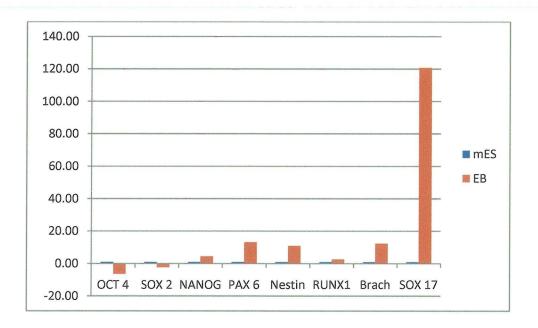


Figure 3. Gene expression following EB differentiation. Fold difference is shown realitye to undifferentiated iPS cell line.

	OCT 4	SOX 2	NANOG	PAX 6	Nestin	RUNX1	Brach	AFP	SOX 17
mES	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00	1.00
EB	-6.35	-2.33	4.47	13.11	10.97	2.66	12.35	1264521.09	120.64

Table 1. Fold difference values of gene expression of EB. Fold difference is shown as fold difference to undifferentiated cells.

#### **Transgene Induction**

Briefly, cells are plated on a gelatinized 6-well plate at low density for 3 days and maintained in medium containing both puromycin and doxycycline. On day 3, transgene expression is induced by withdrawal of doxycycline. After 48 hours, the cells are harvested for RNA extraction, followed by quantitative PCR using specific primers targeting the tansgene (SYBR green PCR Master Mix, ABI) Amplification results are normalized to the histone H2A transcript and analyzed using the delta-delta Ct method to approximate fold change in gene expression (induced to uninduced control vector).

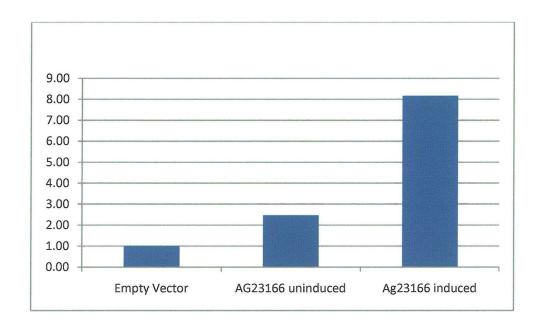


Figure 4. Gene expression following transgene induction. Fold difference is shown relative to uninduced mES cell line containing empty vector.

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